AMENDMENTS TO THE SPECIFICATION

Please replace the paragraph beginning at page 19, lines 20-26 with the following amended paragraph:

Figure 6 shows how an *Xba*I site can be made sensitive to methylation.

The RE cuts at the sites shown by arrows. The boxed sequence is methylated in a dam^+ strain thereby altering the *Xba*I recognition site. The sequence however is not methylated in a dam^- strain, and so can still be cleaved by *Xba*I. The *Xba*I recognition sequence (5'**TCTAGA**3') can therefore be selectively cleaved by *Xba*I I. Assembly of DNA units uses only one restriction enzyme - *Xba*I. Figure 6 discloses SEQ ID NOS:1-2, respectively, in order of appearance.

Please replace the paragraph beginning at page 20, lines 19-24 with the following amended paragraph:

Figure 13 shows the amino acid sequence alignment of the recA protein of S. lividans (S.l.) (SEQ ID NO:3) and S. ambofaciens (S.a.) (SEQ ID NO:4). Percent similarity: 96.496, percent identity: 95.418. Match display thresholds for the alignment(s):

I = identity

: = 2

. = 1

Please replace the paragraph beginning at page 20, lines 26-28 with the following amended paragraph:

Figures 14A and 14B show a DNA sequence alignment of the recA gene S. lividans (S.l) SEQ ID NO:5) and S. ambofaciens (S.a) SEQ ID NO:6). Start of the gene is from 'ATG' and stop is 'TGA'. Percent similarity: 94.713, percent identity: 94. 713.

Please replace the paragraph beginning at page 25, lines 9-16 with the following amended paragraph:

A similar system to the *XbaI/dam* system described above, uses the restriction enzyme *Fok*I which has the recognition site:

5' GGATG(N)$_9$ 3' (SEQ ID NO:7)

3' CCTAC(N)$_{13}$ 5' (SEQ ID NO:8)

with the *dcm* methylase of *E coli*. Adding CCA or CCT to the 5'end of the *Fok*I recognition site would make the site dcm sensitive. Furthermore, if the sequence TCTAGA were inserted into the redundant section of the *Fok*I restriction site, then the enzyme could be used to generate '*Xba*I-cut ends'.

Please replace the paragraph beginning at page 25, line 20, through page 27, lines 1-10 with the following amended paragraph:

Construction of the final expression plasmid pAR1 0 was carried out in several steps, as follows. The ten PKS DNA units were amplified by PCR using *pfu* DNA polymerase. The respective regions of *eryAI* gene, as well as the oligonucleotides used for each PCR are outlined:
LM - segment of *eryAI* gene (Bevitt *et al.*, 1992) extending from nucleotide (N) 588 to N 2389;
5'GGC**ATATG**GCGGACCTGTCAAAGCTCTCCGACAGT3' (SEQ OD NO:9) and
5'GG**TCTAGATC**CCAGCCGCGGTCGGTCGGCAGTCCCG3' (SEQ ID NO:10);
KS1 - segment of *eryAI* gene extending from N 2384 to N 3769;
5'GG**TCTAGA**CTCGCTGTTCCACCCCGACCCCACGCGCTCGGGCACCGCGCACCA3'
SEQ ID NO:11) and
5'GG**TCTAGATC**GCGCAGCGCGGCGGACTCGTCGACGGGGGCGAAGGCGG3' (SEQ ID NO:12),

AT1 - segment of *eryAI* gene extending from N 3764 to N 4813;
5'GG**TCTAGA**CGGTCTCGCGACGGGAAACGCCGACGGTGCCGCCGTTGGAA3' (SEQ ID NO:13)

and

5'GG**TCTAGATC**CACCGCGACACCGGCGGCGAACGCGCGGGAGAGCGCTTCGC3'

(SEQ ID NO:14),

KR1 - segment of *eryAI* gene extending from N 4808 to N 6316;

5'GG**TCTAGA**GTCGGTGCACCTGGGCACCGGAGCACGCCGGGTGCCCTT3' SEQ ID NO:15)

and

5'GG**TCTAGAT**CGTCGAAGAGCCTGGTCGGGCGCTGCGCGGTGTA3' SEQ ID NO:16),

ACP 1 - segment of *eryAI* gene extending from N 6311 to N 6679;

5'GG**TCTAG**ACGACGCGCGGCGGGCTGCGCCGCAGGCGCCGGCCGAACCGCGGG3' (SEQ ID NO17)

and

5'GG**TCTAGATC**GGCCGTGGTCGCCGGTGCCGCCTGCTCGGCT3' SEQ ID NO:18),

KS2 - segment of *eryAI* gene extending from N 6674 to N 8200;

5'GG**TCTAGA**CGAGCCGATCGCGATCGTCGGCATGGCGTGCCGGCTGC3' (SEQ ID NO:19)

and

5'GG**TCTAGATC**GTGCACGGCCTCGGCGGTGTCGGCGGCGAGCACCGCGGCCCGCTCCTC3' (SEQ ID NO:20),

AT2 - segment of *eryAI* gene extending from N 8195 to N 9340;

5'GG**TCTAGA**GGCGGTGGCCGACGGCGCGGTGGTT3' (SEQ ID NO:21)

and

5'GG**TCTAGATC**GTCAC GAG G G GTG GTG CG GTCCG GCAG CAG CCAGAA3' (SEQ ID NO:22),

KR2 - segment of *eryAI* gene extending from N 9335 to N 10639;

5'GG**TCTAGA**CGGCTGGTTCTACCGGGTCGACTGGACCGAG3' (SEQ ID NO:23)

and

5'GGT**CTAGATC**CGGCCGGGGCCGGGCGGCGGTGTAGGACT3' (SEQ ID NO:24),

ACP2 - segment of *eryAI* gene extending from N 10634 to N 10966;
5'GGTCTAGACCGCATCGTCACGACCGCGCCGAGCGA3' (SEQ ID NO:25) and
5'GGTCTAGATCGGCGTCGAGGAAA3' (SEQ ID NO:26),

TE - segment of e *eryAIII* gene (Donadio *et a*l. 1991) extending from N 8753 to N 9602;
5'GG**TCTAGA**CAGCGGGACTCCCGCCCGGGAAGCG3' (SEQ ID NO:27) and
5'GGGCTAGC**TCTAGATC**ATGAATTCCCTCCGCCCAGCCAGGCGTC3' (SEQ ID NO:28).

Please replace the paragraph beginning at page 39, lines 1-20 with the following amended paragraph:

The oligonucleotides:

5'- GGTCTAGAATTCGGCAAGGGCGCCGGTCATGCGCAT-3' (SEQ ID NO29) and 5'- GG TCTAGA TGTGCGGCGTCGGCCGGGGCGGCGGAGGCG-3' (SEQ ID NO:30) were used as the forward and reverse primers respectively and the 1000 bp internal region of *S. lividans recA* gene (Nussbaumer and Wohlleben, 1994) was amplified using pfu polymerase. An additional nucleotide (B) was incorporated into the forward primer to generate a frame shift in the amplified *recA* gene fragment. The PCR product was cloned in pUC-18 vector and sequenced to detect for possible errors during PCR. The 1.0 kbp *rec*A fragment, flanked at both ends by an *Xba*I site was then inserted in the expression vector pCJR24 that has a unique *Xba*I site. The ligation mixture was used to transform *E. coli* DH10B cells and the desired plasmid DNA isolated. The resulting plasmid (p*ARec*A24) contains a non- methylated *Xba*I site at the 5' end of the *rec*A gene fragment. The ten PKS DNA units, namely, TE, two each of ACP1, KR1, AT1 & KS1, and LM were inserted into the plasmid pARecA24 to finally yield the expression plasmid p*Rec*AD1TE. This plasmid was used to transform wild-type *S. lividans* protoplasts, and thiostrepton resistant colonies were grown in defined liquid media as described above. The compound (Figure 12) was isolated from the bacterial broth and chemically characterised.